CACNA1B mutation is linked to unique Myoclonus-Dystonia syndrome

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ABSTRACT

Using exome sequencing and linkage analysis in a 3-generation family with an unique dominant Myoclonus-Dystonia-like syndrome with cardiac arrhythmias we identified a mutation in the CACNA1B gene, coding for neuronal voltage-gated calcium channels Ca\(_{v}\)2.2. This mutation (c.4166G>A;p.Arg1389His) is a disruptive missense mutation in the outer region of the ion pore. The functional consequences of the identified mutation was studied using whole cell and single channel patch recordings. High resolution analyses at the single channel level showed that, when open, R1389H Ca\(_{v}\)2.2 channels carried less current compared to WT channels. Other biophysical channel properties were unaltered in R1389H channels including ion selectivity, voltage-dependent activation or voltage-dependent inactivation. Ca\(_{v}\)2.2 channels regulate transmitter release at inhibitory and excitatory synapses. Functional changes could be consistent with a gain-of-function causing the observed hyperexcitability characteristic of this unique Myoclonus-Dystonia-like syndrome associated with cardiac arrhythmias.
INTRODUCTION

Myoclonus-Dystonia syndrome (M-D, MIM #159900) is a rare hyperkinetic movement disorder with myoclonic symptoms (sudden, shock-like movements) combined with dystonia, mainly of the upper part of the body.(1) Psychiatric comorbidity has been described and symptoms often respond to alcohol ingestion.(2-4) Up to 50% of individuals with definite M-D have mutations in the epsilon-sarcoglycan gene (SGCE, DYT11, MIM159900).(5-7) Previously, we described a SGCE-negative 3-generation pedigree with an unique familial M-D syndrome of unknown cause.(8) Affected members of this family (n = 5) suffered from cervical and axial dystonia at rest, writer’s cramp and action-induced foot dystonia. Myoclonic jerks with on EMG semirhythmic bursts of 40–200ms occurred in legs and arms during rest. Additional symptoms, not previously described in M-D, include high-frequency continuous myoclonus in the legs while standing causing unsteadiness. Intermuscular and corticomuscular coherences around 12 Hz were present and the phase differences suggested an afferent (sensory) cortical input. Eye movement recordings suggest cerebellar pathology. Also present and not previously linked to M-D were cardiac arrhythmias and attacks of painful cramps in upper and lower limbs in 3 of 5 affected family members. By combining exome sequencing with linkage mapping in this pedigree we identified three mutations, but focused on a mutation R1389H (rs184841813) in the CACNA1B gene that codes for a presynaptic neuronal voltage-gated calcium channels Cav2.2 (MIM 601012). By studying single channel currents of mutant and wild-type human Cav2.2 channels in a mammalian cell line, we reveal that the R1389H mutation affects the rate of ion flow through single ion channels. These functional analyses of cloned Cav2.2 channels are consistent with a gain-of-function phenotype.

RESULTS

Patients

We used a 3-generation pedigree for our studies, including 5 of 16 family members suffering from an M-D-plus syndrome previously described.(8)(Fig 1) Features compatible with the known M-D phenotype were cervical and axial dystonia at rest, writer’s cramp and action-induced foot dystonia with walking, myoclonic jerks in legs and arms, increasing with action, a positive effect of alcohol on
the jerks, and complaints of hyperventilation or panic attacks. However, new prominent features in this pedigree are the high frequency continuous myoclonus in the legs while standing, causing unsteadiness. Also in 3 of 5 affected family members cardiac arrhythmias and attacks of painful cramps in upper and lower limbs were present. (8) Recently, the single brother (III-4) presented with a sudden onset, fixed dystonic posture of the left ankle and wrist, consistent with functional dystonia. No myoclonic jerks or orthostatic myoclonus were noticed. His diagnosis was confirmed by an independent neurologist. This individual was therefore considered unaffected in further study.

**CACNA1B R1389H**

Linkage analysis assuming a dominant mode of inheritance identified a region of interest of approximately 150 MB. In this region 5 rare heterozygous missense SNVs were found by exome sequencing in affected family member III-2, confirmed by Sanger sequencing. Segregation with disease within the family was seen for three rare SNVs: CACNA1B c.4166G>A:p.R1389H, rs184841813 (NM_000718), VPS13D c.10355A>G:p.Q3452R (NM_015378) and SPTAN1 c.5308C>T:p.R1770C, rs371055930 (NM_001130438). These SNVs were not present in 760 (1012 for CACNA1B) healthy Dutch controls from the Dutch Blood Bank. Both the CACNA1B and SPTAN1 SNVs are present at very low frequencies in the ±6500 exome variant server (EVS, http://evs.gs.washington.edu/EVS/) (EVS MAF 9.2*10^{-4} and 7.6*10^{-5} respectively). Since M-D is a relatively frequent disorder and EVS participants were not screened for movement disorders, the presence of these variants in EVS is not an argument against their pathogenicity.

The VPS13D Q3452R substitution is predicted to be tolerated (SIFT, PolyPhen) as the amino acid is moderately conserved and there is only a small physiochemical difference between Gln and Arg. The R1770C variant in SPTAN1 and R1389H in CACNA1B are predicted to be deleterious and residues are well conserved among species. The lack of an association of previously found mutations in SPTAN1 to M-D, lead us to focus on the CACNA1B variant. The clinical presentation of myoclonic jerks and dystonia in combination with arrhythmias in this family pointed to a possible channelopathy. The CACNA1B gene encodes Ca\(_{\text{v}}\)2.2, the main of pore-forming alpha-1 subunit of the multi-subunit
voltage-gated calcium channel complex that underlies N-type currents in neurons. The preferential location of \( \text{CaV}_2.2 \) is at nerve terminals and they play a critical role in controlling transmitter release at many synapses. (9) Previously identified pathogenic \textit{CACNA1A} mutations in the homologous region of the gene cause familial hemiplegic migraine (V1457L, OMIM 601011.0019) (10) and causing episodic ataxia (type 2) (F1406C, OMIM 601011.0022). (11,12) \( \text{CaV}_2.2 \) null mice have a major locomotion phenotype, showing significantly more activity (our work, unpublished, and (13,14)).

The region of \textit{CACNA1B} coding for the IIIS5 through IIIS6 was Sanger sequenced in 47 patients with an M-D phenotype. All contained wild-type R1389 and no other novel missense SNVs were identified. Given the location of R1389 in the extracellular ion pore loop of S5-S6 in CaV2.2 (Fig 3A), we were able to monitor protein function directly.

**Functional analyses R1389H hCa\(_{\gamma}2.2\)**

To assess the functional consequences of the R1389H mutation on the biophysical properties of Ca\(_{\gamma}\) channels we expressed wild-type and mutant hCa\(_{\gamma}2.2\) channels in the mammalian tsA201 cell line and compared calcium channel currents using whole cell and single channel patch recordings (Fig 2 - 4). Peak and instantaneous current voltage relationships illustrate that currents through wild-type and R1389H mutant channels reverse direction at close to the same voltages (54 mV and 52 mV; Fig 2C, 2F). This suggests that R1389H has little or no influence on ion selectivity. Channel properties that reflect gating mechanisms such as the voltage-dependence of activation and inactivation were also not different between wild-type and R1389H channels (Fig 2D, 2E). Thus the biophysical properties of whole cell currents were not different, although average whole cell hCa\(_{\gamma}2.2\) current densities in cells expressing R1389H were larger compared to wild-type (Fig 2A - 2C). The functional significance of such a difference in current densities in a non-neuronal overexpression system is not clear. The clones are expressed at high level and several factors contribute to overall current density that are independent of the biophysical properties of the channel but depend on cell-type and subcellular location, including translation efficiency, plasma membrane trafficking, protein stability. (15)
We therefore used high-resolution cell-attached patch recordings to make exact measurements of ion flow through single hCa\textsubscript{V}2.2 wild-type and R1389H mutant ion channels (Fig 3, 4). Single channel currents of R1389H were smaller on average compared to wild-type measured under identical conditions (Fig 3B - 3D). At +20 mV, the dominant current amplitude of single wild-type channels was 1.03 ± 0.02 (n = 8 control patch recordings) compared to 0.77 ± 0.01 for R1389H (n = 8 patch recordings; p = 9.1 × 10^{-10}, Student’ s t-test) (Fig 3B - 3D). Further analysis revealed that human Ca\textsubscript{V}2.2 WT channels transition to two different open states the larger of which, O\textsubscript{2} (1.03 pA), dominates and represents about 85% of all openings, compared to O\textsubscript{1} (0.82 ± 0.02 pA, 8 patches) (Fig 3B, 4A). The presence of two different open states in WT recordings is shown as an excess of smaller amplitudes in the amplitude histogram calculated from >500 openings of more than 0.3 ms duration (Fig 4A). We observed direct transitions between O\textsubscript{1} and O\textsubscript{2} within a single opening (Fig 4B), suggesting that individual Ca\textsubscript{V}2.2 channels can exist in two open states that conduct ions at different rates. By contrast, single R1389H channels open predominantly to amplitudes that are on average not significantly different from the O\textsubscript{1} amplitude of WT channels (0.77 ± 0.01 pA, n = 8 patches; p = 0.35, Student’ s t- test) (Fig 4C, 4D). Our data suggest that R1389H stabilizes the smaller amplitude open state as compared to WT.

DISCUSSION

We report a missense mutation (c.4166 G>A; p.Arg1389His, rs184841813) in the CACNA1B gene in a 3-generation family with a Myoclonus-Dystonia-like syndrome characterized by progressive myoclonic jerks, dystonia and an action-induced lower limb myoclonus combined with cardiac arrhythmias in one pedigree. Clinical and electrophysiological assessment of patients from this family is most consistent with a subcortical origin with possible cerebellar involvement. CACNA1B codes for the pore-forming subunit of presynaptic Ca\textsubscript{V}2.2 channels (MIM 601012) that together with Ca\textsubscript{V}2.1 (CACNA1A), control depolarization-induced calcium entry and transmitter release from the majority of presynaptic terminals at mammalian synapses. Thus a gain of function that influences current flow through the Ca\textsubscript{V}2.2 channel could directly impact synaptic transmission and could explain the hyperexcitability, characteristic of Myoclonus Dystonia syndrome.
After finalizing our studies, the considered unaffected sibling (III-4) presented with a sudden onset, fixed dystonic posture of the left ankle and wrist without myoclonic jerks. His presentation was very different from the affected members and consistent with functional dystonia. We therefore, still consider him unaffected. His clinical diagnosis was confirmed by an independent neurologist. He did not carry the variant and did not share the same haplotype as affected individuals. This twist in the story illustrates the difficulty of family studies in dystonia.

Ca_{v}2.2 forms a complex with Ca_{v}a_{2}d and Ca_{v}b subunits that localizes to active zones of presynaptic nerve terminals throughout the brain and nervous system. Together with Ca_{v}2.1, the activity of Ca_{v}2.2 controls the amount of transmitter released in response to depolarization of the presynaptic terminal. At most synapses, the concerted action of Ca_{v}2.1 and Ca_{v}2.2 channels regulate depolarization-dependent presynaptic calcium entry, while at other synapses presynaptic Ca_{v}2.1 or Ca_{v}2.2 channels control calcium entry exclusively.(9,16,17) Our functional analyses reveal reduced rate of ion flow through single ion channels compared to wild-type. Other essential biophysical channel properties were unaltered in R1389H channels including ion selectivity, voltage-dependent activation or voltage-dependent inactivation. The activation duration of single R1389H mutant channels tended to be longer on average, compared to WT channels. For example, longer activation times of R1389H compared to WT allowed measurements of channel amplitudes at more negative potentials – when channel open times are shorter – and wild-type controls currents were too short to measure accurately (Fig 3C, 3D). A longer activation time at the single channel level could give rise to larger current densities in whole cell recordings of R1389H compared to wild-type controls (Fig 2). We did observe consistent differences in the overall expression level of Ca_{v}2.2 channels, but current density measurements depend on several factors that may be influenced by the specific over-expression system used, cell-type, trafficking to and from, and stability at the plasma membrane. Thus the functional significance of this observation remains an open question. By contrast, single channel current amplitude is an exact measure that is influenced by variables that are readily control in our experiments (concentration of the permeant ion, membrane voltage, and temperature.)
Our study is the first to report high-resolution recordings of single human Cav2.2 channel currents and they reveal the presence of at least two subconductance states that have important disease relevance. R1389H stabilizes Cav2.2 channels in the lower current amplitude open state that is observed much less often in wild-type recordings. Subconductance states are a feature of L-type Cav1.2 channels and these have been attributed to proton block of ion flow.(18-20) Arginine to histidine disease-causing mutations, occur with relatively high frequency. The R1389H exonic mutation in CACNA1B could destabilize potential interactions that depend on the positive charge of arginine at physiological pH such as salt bridge formation between aspartate and glutamate, and could result in sensitivity to changes in the extracellular pH. For example, replacement of a highly conserved arginine in the ion pore of an inwardly rectifying potassium channel with histidine, reduces the flow of ions through single channels.(21)

Mutations in the closely related CACNA1A gene are linked to several autosomal dominant neurologic disorders including familial hemiplegic migraine (FHM), episodic ataxia type 2 (EA-2), and spinocerebellar ataxia type 6 (SCA-6). Notably, one of the FHM-causing mutations (V1457L)(10) and a mutation causing EA-2 (F1406C) (11,12) occur in the S5-S6 loop of the CACNA1A gene and are in a position homologous to R1389H of CACNA1B. The V1457L Cav2.1 mutant channel shows lower unitary current amplitudes compared to wild-type currents, as well as altered channel gating.(22)

Surprisingly, mutations in CACNA1B have not been demonstrably linked to any human hereditary disorder previously. Common variants in CACNA1B are associated with cerebral infarction(23), bipolar disorders and schizophrenia.(24,25) Here we provide evidence that a disruptive missense mutation in the outer region of the ion pore of Cav2.2 channels may cause an unique M-D syndrome associated with cardiac arrhythmias. Ziconotide (ω-conotoxin MVIIA) is a selective Cav2.2 channel inhibitor used clinically to relieve otherwise intractable chronic pain and might be a good candidate for therapeutic intervention in our family. By discovering new genes in dominant forms of dystonia we
gain more insight in the pathophysiology of dystonia and hopefully discover promising pharmaceutical therapies for certain forms of (myoclonus) dystonia.

MATERIALS AND METHODS

Patients

All patients were previously described. The study was approved by the Medical Ethical Committee of the Academic Medical Center Amsterdam. All patients were examined by a movement disorder specialist (MAJT). Investigations were performed with informed consent.

We performed linkage analysis in this three generation family and combined these results with the exome data, obtained in the index case.

Linkage analysis

Linkage analysis was performed using the Affymetrix GeneChip Human Mapping 250K Nsp Array in 5 affected and one unaffected sibling from 3 generations. The linkage analysis was performed in family members II-2, III-2, III-3, III-4, III-6, IV-6 (see Fig 1). There are four informative meiosis in this panel. For a dominant disorder 0.3 LOD-score points can be obtained per meiosis, providing a maximum LOD score of 1.2. Linkage analysis was performed with the program ALLEGRO making use of a reduced marker panel of approximately 20000 SNPs. Autosomal dominant mode of inheritance with full penetrance was assumed with a disease allele frequency of 0.0001. For the 13 linked regions, again calculations with ALLEGRO was performed with all SNPs from 250K-Chip to determine more accurately the limiting SNPs of the linkage regions.

Exome sequencing

Exome sequencing was performed using enriched libraries (NimbleGen SeqCap EZ Exome v2 Sequence Capture beads) sequenced on a SOLiD v4 platform (Life Technologies), which created paired end reads of 50 bases forward and 35 bases reverse. Colour space reads were mapped to the reference human genome (hg18) with SOLiD Bioscope software version 1.3 (Life Technologies). Average coverage of the exome sequencing was 91% for the target and average depth was 58x after
removal of duplicate reads. PCR duplicates were removed prior to variant calling using MarkDuplicates from the Picard software (picard.sourceforge.net/index.shtml). Reads were realigned around known ‘indels’ with the Genome Analysis Toolkit (Broad Institute). Variants were called using the GATK’s Unified Genotyper. GATK’s VariantAnnotator was used to annotate variant calls in preparation of filtering. Non-synonymous variants in coding regions that are not present in dbSNP 130 were identified with ANNOVAR.(26) SNV population frequency was examined in 760 controls from the Dutch Blood Bank with TaqMan Custom Endpoint Genotyping assays (Applied Biosystems, Foster City, USA). Variants were confirmed by Sanger sequencing.

**Electrophysiological analyses of human-derived Ca\textsubscript{v}2.2 clones**

Full-length human Ca\textsubscript{v}2.2 cDNA (ORF 7094 bp, pSAD442-1) was cloned from human brain and spinal cord and inserted into pcDNA6 mammalian expression vector. pSAD442-1 contains alternate exons \([+e10a, +18a, \Delta19a, +e31a, +e37b, +e46]\).(27) The R1389H mutant was generated using standard mutagenesis methods and sequence confirmed. Wild-type and mutant hCa\textsubscript{v}2.2 were transiently expressed in human tsA201 cells with Ca\textsubscript{v}\(\beta\)\textsubscript{3}, Ca\textsubscript{v}\(\alpha\)\textsubscript{2}\(\delta\)-1 and enhanced green fluorescent protein cDNAs (eGFP; BD Bioscience) as described previously.(28) Whole cell and single calcium channel currents were recorded 24 hours after transfection. cDNA concentrations were measured and visualized in agarose gels to ensure these were not different between mutant and wild-type transfections. Recording from wild-type and mutant Ca\textsubscript{v}2.2 clones were interspersed and the experimenter blind to clone identity during recording and analysis. Macroscopic, whole cell recordings were obtained from 7 separate transfections for both WT and R1389H cDNAs. The identity of WT and R1389H clones were revealed post analysis. Standard whole cell patch clamp and single channel recordings were used to compare calcium channel currents.(15,28) External solution for whole cell recordings contained: 1 mM CaCl\textsubscript{2}, 4 mM MgCl\textsubscript{2}, 10 mM HEPES, 135 mM tetraethylammonium (TEA) chloride, pH adjusted to 7.2 with TEAOH and internal solution contained: 126 mM CsCl, 10 mM EGTA, 1 mM EDTA, 10 mM HEPES, 4 mM MgATP, pH 7.2 with CsOH. Whole-cell calcium currents were evoked by voltage-steps and currents leak subtracted on-line using a P/-4 protocol. Data
were sampled at 20 kHz and filtered at 2 kHz (-3 dB) using an Axopatch 200A amplifier (Molecular Devices, LLC). For single channel recording, external solution contained: 110 mM BaCl₂, 2 CsCl, 10 HEPES, pH adjusted to 7.2 with BaOH and bath solution contained: 135 potassium aspartate, 10 EGTA, 5 HEPES, 5 KCl, pH adjusted to 7.2 with KOH. Electrodes were Sylgard® 184 (Dow Corning, Co) coated and had resistance of 5-8 MΩ. Single channel data were sampled at 10-20 kHz and filtered at 2 kHz (-3 dB) using Axopatch 200B (Molecular Devices, LLC). Clampfit10 (Molecular Devices, LLC) was used to classify and analyze single-channel events. Leak subtraction was performed off-line by best-fit curve to null traces. All recordings were obtained at room temperature.

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CONFLICT OF INTERESTS STATEMENT

The authors report no conflict of interest.
REFERENCE


LEGENDS TO FIGURES

Figure 1. Pedigree of M-D family with R1389H mutation in CACNA1B. Unaffected and affected family members are indicated by clear symbols and blackened symbols, respectively. Diagonal bars through the symbols denote deceased individuals. * denote examined and included in linkage studies. In individual III-2 exome sequencing was performed.

Figure 2. Whole cell calcium currents recorded from tsA201 cells transiently expressing human R1389H and wild-type control (WT) C\textsubscript{a}\textsubscript{v}2.2 channels. R1389H or WT C\textsubscript{a}\textsubscript{v}2.2 cDNAs were expressed together with C\textsubscript{a}\textsubscript{v}\beta\textsubscript{3}, C\textsubscript{a}\textsubscript{v}\alpha\textsubscript{2}δ-1 and enhanced green fluorescent protein (eGFP). Currents were recorded from fluorescent cells 24 hours after transfection using 1 mM calcium as the charge carrier. A, Current traces recorded from cells expressing R1389H or WT C\textsubscript{a}\textsubscript{v}2.2. B, Peak current densities at +10 mV shown together with average values for R1389H: 97.9 ± 15.0 pA/pF (21 cells) and WT: 55.1 ± 9.1 pA/pF (26 cells) (p = 0.014, Student’s t-test). C, Average, peak current-voltage relationships for R1389H and WT. Currents were evoked by depolarizing steps to indicated potentials from a holding potential of -80 mV. Boltzmann-linear functions were fit to individual data sets to estimate average activation mid-point (V\textsubscript{1/2}), slope factor (k) and reversal potential (V\textsubscript{rev}). Averages ± SE were for R1389H (n = 21) and WT (n = 26), V\textsubscript{1/2}: -0.18 ± 0.98 mV and 0.85 ± 1.03 mV (p = 0.47); k: 4.8 ± 0.2 mV and 5.8 ± 0.5 mV (p = 0.06); and V\textsubscript{rev}: 51.6 ± 1.2 mV and 54.2 ± 0.8 mV (p = 0.08). Only currents > 40 pA at 10 mV were included in the analysis (~75% of fluorescent cells for R1389H and WT). D, Activation curves measured from tail current amplitudes at -60 mV expressed as I/I\textsubscript{max} for a range of test potentials. Boltzmann functions were fit to estimate V\textsubscript{1/2} and k. Average ± SE for R1389H (n = 21) and WT (n = 26) were for V\textsubscript{1/2}: 12.6 ± 1.1 mV and 13.6 ± 0.9 mV (p = 0.47) and k: 8.4 ± 0.8 mV and 8.3 ± 0.5 mV (p = 0.88). E, Inactivation curves measured as fraction of current available in response to a voltage step to 10 mV from different pre-pulse potentials. Boltzmann functions fit to each data set were used to estimate average ± SE values for R1389H (n = 11) and WT (n = 9) for V\textsubscript{1/2}: -48.0 ± 1.8 mV and -51.4 ± 3.8 mV (p = 0.40) and k: 10.7 ± 0.3 mV and 11.6 ± 0.4 mV (p = 0.09). F, Instantaneous current-voltage relationship represents the total current flow through all open channels, the data are well fit by a straight line.
Figure 3. Comparison of current flow through single R1389H and WT control Ca\textsubscript{v}2.2 channels recorded in cell-attached patches. A, Location of R1389H in the outer region of the Ca\textsubscript{v}2.2 ion pore (red star) and two calcium ions depicted in the narrow part of the selectivity filter (brown circles).\( ^{(29,30)} \) B, single channel current amplitudes measured from cell-attached patch recordings of R1389H (red triangle) and WT (black circles and open triangles) Ca\textsubscript{v}2.2 channels. Solid symbols are the average values across patches, and open symbols are averages of 5-100 measurements from channels that were open for long enough to reach full amplitude. WT channels opened to two levels \( O_1 \) and \( O_2 \), but openings to \( O_2 \) dominate (see Fig. 4). Single channel conductances were between 23 pS and 27 pS. Examples of recordings of WT (C) and R1389H (D) channel currents evoked by step depolarizations to +20 mV, +10 mV, and 0 mV. Dotted lines indicate average single channel current amplitudes at different membrane potentials shown in B.

Figure 4. Comparison of single channel currents in cell-attached patch recordings from WT and R1389H. Amplitude histograms calculated from single channel analysis of at least 1,000 events per cell-attach patch recording of WT (A; 1,033 events) and R1389H (C; 3,256 events) channels at +20 mV. Channel open and closed transitions were fit to each event that were longer than 0.3 ms (more than 3 points). Amplitude histograms of closed (C) and open (O) states were fit with one or the sum of two Gaussian functions. A, \( O_1 \): \( \mu_1 = 0.70 \text{ pA} \) and \( A_1 = 2.95 \); \( O_2 \): \( \mu_2 = 0.93 \text{ pA} \) and \( A_2 = 17.29 \). C,(B) \( \mu = 0.67 \text{ pA} \) and \( A = 59.5 \). Examples of single channel current are shown for WT (B) and R1389H (D) channels on two different time scales. WT Ca\textsubscript{v}2.2 channels can open from the closed state to two different open levels, \( O_1 \) and \( O_2 \), and transition directly between \( O_1 \) and \( O_2 \) without closing. R1389H channels opened to a single level, \( O_1 \). Dotted lines indicate average current amplitudes from Gaussian fits (A, C).
ABBREVIATIONS

EA-2 - episodic ataxia type 2
EMG – electromyography
FHM - familial hemiplegic migraine
MAF – minor allele frequency
M-D – Myclonus-Dystonia
SCA-6 - spinocerebellar ataxia type 6
SGCE - epsilon-sarcoglycan
SNV – single nucleotide variant
WT – wild type